

Marine Turtle Newsletter

Number 62 July 1993

Editors:

*Karen L. Eckert & Scott A. Eckert
Hubbs-Sea World Research Institute
1700 South Shores Road
San Diego, California
92109 USA*

Editorial Board:

*Nat B. Frazer
Nicholas Mrosovsky
David W. Owens
Peter C. H. Pritchard
James I. Richardson*

time consultant, continuing to do many of the same things he did before his retirement. His hobbies include hunting, fishing, gardening, and, in all likelihood, giving grief where it is richly deserved. We all miss you Jack Woody.

MARYDELE DONNELLY, RD 4, Box 4427, Stroudsburg, Pennsylvania 18360 USA.

Editor's Note: It's a difficult assignment to capsule the career of one as accomplished and as admired as Jack Woody, and we sincerely thank Marydele for accepting the challenge! And on behalf of a multitude of friends and fans who could not be included in this brief *adieu*, we wish you, Jack, all good things and many very, very happy years.

FWS SELECTS NEW SEA TURTLE COORDINATOR

Dr. Richard Byles (FWS Division of Endangered Species, P.O. Box 1306, Albuquerque, New Mexico 97103; Tel: (505) 766-3972, FAX: (505) 766-8063) has been selected for the position of the U.S. Fish and Wildlife Service (FWS) National Sea Turtle Coordinator. Byles replaces Jack Woody, who retired from Federal service this year and had held the post since it was created. Byles brings to the job 20 years of experience in the conservation and scientific study of sea turtles, and extensive dealings with Latin American and U.S. sea turtle researchers, government agencies, and non-government organizations. He has served on the Kemp's Ridley Recovery Team and the East Pacific Sea Turtle Recovery Team and is editing the status reviews of all U.S. sea turtle species. Byles is a member of the IUCN/SSC Marine Turtle Specialist Group, is an adjunct professor in biology at the University of New Mexico, and is a temporary member of the graduate faculty of Texas A&M University. Byles has spent the last seven years as the FWS Sea Turtle Biologist, working closely with Woody. His expertise and contacts will certainly aid us in achieving Service goals for sea turtles and their habitat. Source: excerpted from a U.S. Department of Interior Memo, 28 April 1993; Lynn B. Starnes, Acting Regional Director.

RESUSCITATION OF SEA TURTLES

Several techniques have been described for resuscitation of comatose sea turtles. These include (a) electrical stimulation of the pectoral region (Shoop, 1982), (b) periodic compression (pumping) of the plastron with the turtle in a supine position (Hopkins and Richardson, 1984), and (c) insertion of a plastic tube into the trachea followed by gentle blowing into the tube at irregular intervals (Balazs, 1986). Given the potential for adverse effects, electrical shock cannot be recommended for routine resuscitation of sea turtles. Pumping of the plastron is also a questionable resuscitation technique. Balazs (1986) reported that plastral pumping does not ventilate the lungs with air because of glottal lock (i.e., airway closure). In addition, placing turtles in a supine position during plastral pumping causes the viscera to compress the dorsally located lungs, thereby reducing lung volume and hindering lung inflation. We describe herein a modification of the resuscitation technique of Balazs (1986) that has been used successfully with Kemp's ridley sea turtles (*Lepidochelys kempi*) recovering from general anesthesia and anoxic submergence.

The well-accepted ABC's of resuscitation are to preserve (a) a clear Airway, (b) normal Breathing, and (c) Cardiac function. In our experience, airway closure resulting from glottal lock and cessation of normal ventilatory patterns (rather than cardiac dysfunction) limit the

recovery of comatose sea turtles. Nevertheless, inadequate gas exchange will eventually compromise normal cardiac function and decrease the survival potential of comatose turtles. Therefore, the present technique was designed to resuscitate the turtle by manually or mechanically increasing pulmonary gas exchange during the comatose period. In our procedure, the turtle is maintained in a prone position out of water. A mouth gag is used to hold the jaws open and permit easy access to the glottis, which is located posterior to the tongue on the floor of the mouth. Ideally, the mouth gag should be constructed from wood or polyvinylchloride (PVC) in order to minimize damage to the keratinous sheaths that cover the jaws. The turtle is intubated with an appropriately-sized endotracheal tube with a low pressure cuff. Endotracheal tubes that are too large damage the tracheal mucosa, whereas tubes that are too small increase the resistance of gas flowing through the tube and decrease effective respiratory resuscitation. Suitable endotracheal tubes for 4-23 kg Kemp's ridley turtles are 5-10 mm outside diameter (3-7 mm inside diameter). The endotracheal tube is carefully inserted through the glottis into the trachea, to a depth of approximately 5-8 cm. The consequence of inserting the tube past the tracheal bifurcation is inflation of only one lung, and therefore, inadequate gas exchange and possible barotrauma. Once the tube is in position, the low pressure cuff is inflated to prevent air leakage on inspiration. The turtle is then ventilated with air repeatedly and at regular intervals. Ventilation frequency depends on the duration of the comatose period. Minimum ventilation rate should be 2-4 breaths per minute and should be increased the longer the turtle has been comatose.

Lung volumes in reptiles vary as a function of body weight (Tenney and Tenney, 1970). However, the requisite lung volume of individual sea turtles may be unknown at the time of intubation. Generally, the procedure for small turtles is to gently inflate the lungs until the carapace begins to move. Inspiratory duration should be between 1-2 seconds and lung inflation should be slow and gradual throughout the inspiratory period. The preferred method is to underinflate the lungs with less than maximal volumes of air, while augmenting gas exchange by increasing the ventilatory frequency. This ensures that the lungs are not overinflated, which may cause significant and irreparable lung damage. We recommend ventilating the turtle with a hand-held resuscitator bag in cases where mechanical ventilation is unfeasible (i.e., in the field), rather than the technique of blowing into the tube for two reasons. First, human-exhaled gases contain less O₂ (17-18%) and more CO₂ (about 5%) than air (about 21% O₂, 0.03% CO₂). Second, zoonotic diseases may be contracted by the person performing the oral resuscitation procedure.

The low pressure cuff on the endotracheal tube should be deflated when the turtle begins to revive. In our experience with comatose Kemp's ridleys, immediate post-revival characteristics include movement of the pectoral and pelvic muscles, hyperventilation (6 to 10 breaths per minute), and significant increases in heart rate. Heart rate (non-invasive femoral artery ultrasonic Doppler flow probes) and cloacal temperature should be monitored during the entire resuscitation procedure. We have found that revival of comatose turtles is adversely influenced by cloacal temperatures outside the range 25-30 °C. Revival takes longer at temperatures below 25 °C. Temperatures above 30 °C may approach the critical maximum. In all cases, comatose and/or revived turtles should be transported to knowledgeable authorities (e.g., local veterinarian, zoo) for continued treatment and monitoring. Manual ventilation must be continued during transport.

To date, we have found no resuscitation technique that will work with "wet drowned" sea turtles. One 20 kg Kemp's ridley was found to "wet drown" following a period of anoxic submergence. The turtle was conscious and active upon surfacing but quickly became lethargic. After one hour of intermittent unassisted ventilation with no improvement in blood gasses or pH, the turtle ceased ventilating and its heart rate fell from 42 to 6 beats per minute. At that

point, the turtle was intubated and its lungs mechanically ventilated with room (ambient) air. Seawater was collected from the lungs via the intubation tube, providing clear evidence that the turtle had inhaled water and "classically" drowned. Continued mechanical ventilation did not improve blood gases (although heart rate increased six-fold), consistent with the view that the inhaled seawater had irreparably damaged the blood-gas barrier, hindering gas exchange. Unfortunately, the turtle died within 24 hours. Conversely, a comatose Kemp's ridley survived severe anoxic acidosis ("dry drowning" where, as a result of glottal lock, the turtle does not inhale water), but required mechanical ventilation for 48 hours before it would respond to tactile stimulation and ventilate unassisted. The turtle was returned to water and did not demonstrate any adverse side effects from the anoxia or the ventilation procedure.

The technique we have described for resuscitation of comatose sea turtles is easy to perform, can be used efficiently in the laboratory or in the field, and does not require substantial investment in equipment. Endotracheal tubes and manual resuscitators can be purchased for less than US\$ 70 from commercial suppliers of hospital equipment. Cloacal thermometers and ultrasonic Doppler probes are useful but optional equipment.

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ERICH K. STABENAU, National Marine Fisheries Service, Galveston Lab, 4700 Avenue U, Galveston Texas 77551 USA (*), PAULA F. MOON, Cornell University, College of Veterinary Medicine, Ithaca, New York 14853 USA and THOMAS A. HEMING, University of Texas Medical Branch, Departments of Internal Medicine and Physiology and Biophysics, Galveston, Texas 77550 USA. (*) Current address: Dept. Internal Medicine/Pulmonary Division, Route H76, University of Texas Medical Branch, Galveston, Texas 77550 USA.

TAG RETURN FROM A MALE GREEN SEA TURTLE

On 25 July 1989 two adult male green turtles (*Chelonia mydas*) were acquired from local fishermen, tagged and released at Tortuguero, Costa Rica, as part of a feasibility study for a project tagging male green turtles by the Caribbean Conservation Corporation (CCC). The turtles were purchased at sea directly from the fishermen who caught them approximately 2 km offshore of the southern end of the Tortuguero nesting beach in the vicinity of Jalova. The turtles were harpooned on the morning of 25 July by fishermen of the traditional turtle *cayuco* "Kentucky" and were purchased at a price slightly greater than the standard rate for male turtles of this size (3000 and 2650 colones, respectively; about 65 colones=1 US\$). Both turtles were returned by boat to the CCC research station, measured, tagged and released into the sea at the mouth of the Tortuguero River. They measured 94.9 and 84.8 cm straight carapace length and weighed 230 and 185 lb, respectively. Each had an elongated tail of 50.5 and 45.3 cm length, respectively. Each received four model 681 INCONEL tags, one in each flipper, front and rear.